



Puritan TSB Salt Transport Medium

INTENDED USE

Puritan TSB Salt Transport Medium is an enrichment medium used for isolation of *Staphylococcus aureus* spp, particularly methicillin-resistant *Staphylococcus aureus* (MRSA).

SUMMARY AND EXPLANATION

S. aureus is one of the most common causes of skin and soft tissue infection in both the health care and community settings.¹ Enrichment broths are commonly used to increase sensitivity testing for MRSA by increasing isolation rates. Tryptic Soy Broth (TSB) serves as the base medium for Puritan TSB Salt Transport Medium to enhance the growth of *S. aureus*. TSB contains enzymatic digest of casein and enzymatic digest of soybean meal, which provide amino acids and complex nitrogenous compounds that promote microbial growth. Dextrose acts as a carbon energy source that facilitates growth. Dipotassium phosphate acts as a buffering agent. Sodium chloride is added to inhibit or partially inhibit microorganisms other than *S. aureus*.

FORMULATION PER LITER

Tryptic Soy Broth powder
Sodium Chloride
Demineralized Water

pH 7.3 ± 0.2 @ 25°C

PRECAUTIONS

For *In Vitro* Diagnostic Use

- Clinical specimens are considered biohazard and must be handled in a manner to protect laboratory personnel
- To be used by trained and qualified personnel using aseptic technique
- Clinical samples may contain human pathogens including hepatitis virus and Human Immunodeficiency Virus. Institutional and universally recognized guidelines should be followed when handling items contaminated with blood and other body fluids.²
- Specimen vials and other contaminated materials must be sterilized by autoclave before discarding.
- Do not use if the vial is damaged or detected evidence of contamination, discoloration or leakage.

STORAGE

For optimum performance, store at 2-25°C. Avoid freezing and overheating.^{3,4}

Item Number	Product Description	Pack Size
TSB-200	Blue polypropylene screw-cap vial with 2 mL of TSB Salt Medium.	50 / box

LABORATORY SPECIMEN PROCESSING

TSB Salt Transport Medium Collected Sample

1. Vortex the inoculated TSB Salt transport medium for approximately 10 seconds.
2. Incubate inoculated TSB Salt transport medium at 35 ± 2°C.
3. Examine the TSB Salt transport medium for growth after 18-24 hours.
4. Aseptically remove aliquots of the TSB Salt transport medium and inoculate on to an appropriate selective agar plate.

Opti-Swab® Liquid Amies Collected Sample

1. Obtain tubes of TSB Salt Transport Medium and unscrew cap.
2. Vortex the inoculated Opti-Swab Liquid Amies for approximately 10 seconds
3. Unscrew the cap and aseptically transfer the swab from the Opti-Swab Liquid Amies to the TSB Salt Transport Medium using sterile forceps.
4. Replace cap on both Opti-Swab Liquid Amies and TSB Salt Transport Medium
5. Follow the procedures stated above for TSB Salt Transport Medium Collected Sample.

SPECIMEN COLLECTION AND HANDLING

Specimens suitable for culture may be handled using various techniques. For detailed guidance, refer to appropriate references.^{5, 6} Specimens should be obtained before antimicrobial agents have been administered.

QUALITY CONTROL

All batches of Puritan TSB Salt Transport Medium are tested prior to release for pH and further evaluated for their ability to promote growth of the following organisms:

CONTROL	INCUBATION	RESULTS
Methicillin-resistant <i>Staphylococcus aureus</i> ATCC 43300	Aerobic, 48 h @ Room Temperature	Good Recovery
<i>Staphylococcus aureus</i> ATCC 6538	Aerobic, 48 h @ Room Temperature	Good Recovery

LIMITATIONS

Definitive identification of *S. aureus* and MRSA requires additional and/or serological tests. Refer to appropriate reference standards for further instructions.^{5, 6}

REFERENCES

1. National Health and Nutrition Examination Survey (NHANES). 2000. Specimen Collection Procedures Manual.
2. Directive 2000/54/EC of the European Parliament and of the Council of 18 September 2000 on the protection of workers from risk related exposure to biological agents at work. Official Journal of the European Communities. L 262/21–45.
3. Versalovic, J., K.C. Carroll, G. Funke, J.H. Jorgensen, M.L. Landry, D.W. Warnock. 2011. Manual of Clinical Microbiology, 10th ed. American Society for Microbiology. Washington, DC.
4. Miller, J.M. 1996. A guide to specimen management in clinical microbiology. American Society for Microbiology. Washington, DC.
5. Forbes, B.A., D.F. Sahm, A.S. Weissfeld. 2007. Diagnostic Microbiology 12th ed. Mosby. St. Louis, MO.
6. Murray, P.R., E.J. Baron, M.A. Pfaller, F.C. Tenover, R.H. Tenover. 2003. Manual of Clinical Microbiology, 8th ed. American Society for Microbiology. Washington, DC.



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