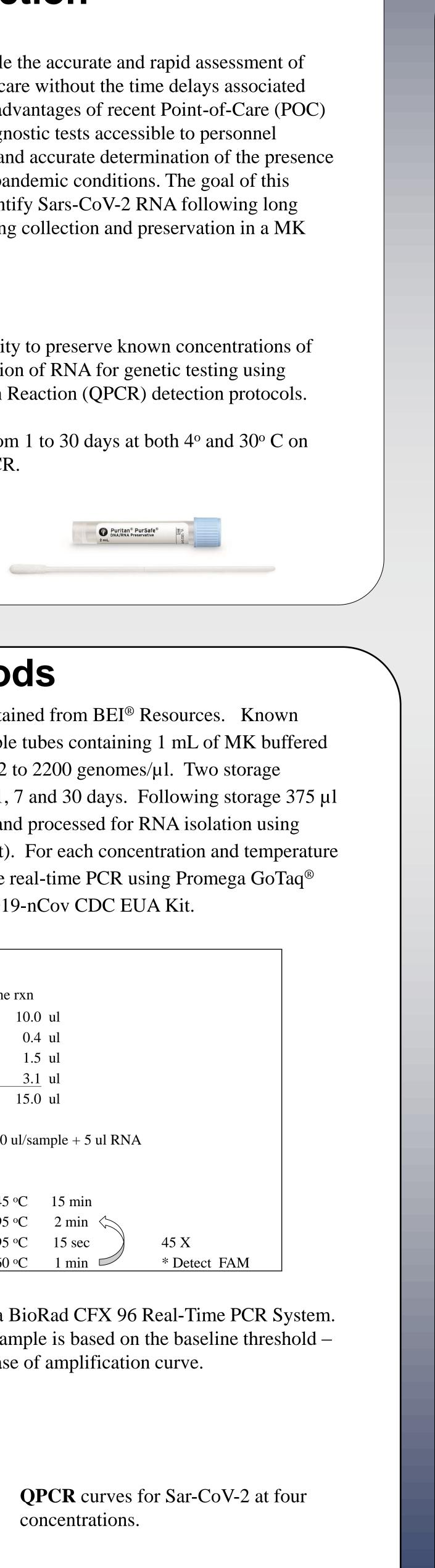


# Introduction

Recent genomic technologies have made possible the accurate and rapid assessment of specific pathogens directly at the site of patient care without the time delays associated with diagnostic laboratories. One of the major advantages of recent Point-of-Care (POC) instrumentation is their ease of use, making diagnostic tests accessible to personnel without specialized laboratory training. Timely and accurate determination of the presence of Sars-CoV-2 is particularly essential in these pandemic conditions. The goal of this study was to assess the ability to detect and quantify Sars-CoV-2 RNA following long term storage in temperatures up to 30°C following collection and preservation in a MK buffered solution.

The goal of this project is

- To evaluate Puritan MK buffer solution's ability to preserve known concentrations of Sars-CoV-2 and allow recovery and quantitation of RNA for genetic testing using standard CDC quantitative Polymerase Chain Reaction (QPCR) detection protocols.
- To assess the effect of storage Sars-CoV-2 from 1 to 30 days at both 4° and 30° C on the RNA concentrations as measured by QPCR.

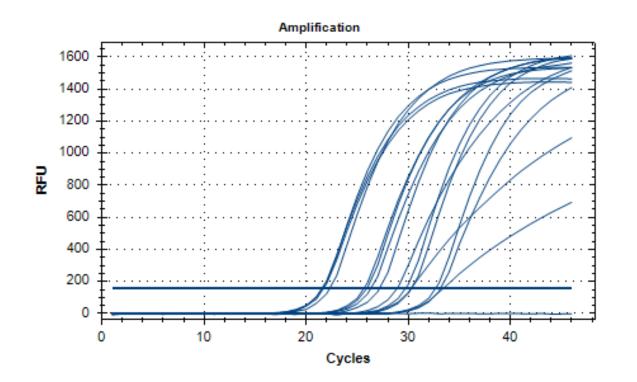


## Methods

Heat-inactivated Sars-CoV-2 (NR-52286) was obtained from BEI<sup>®</sup> Resources. Known concentrations of Sars-CoV-2 were added to sample tubes containing 1 mL of MK buffered solution for final concentrations ranging from 0.22 to 2200 genomes/µl. Two storage temperatures were evaluated, 4° and 30° C for 0, 1, 7 and 30 days. Following storage 375 µl was removed at each time point and temperature and processed for RNA isolation using standard RNA isolation kits (Zymo Viral RNA Kit). For each concentration and temperature three replicates were quantified using Quantitative real-time PCR using Promega GoTaq<sup>®</sup> Probe 1- Step RT-qPCR System and primer set 2019-nCov CDC EUA Kit.

ga GoTAQ Probe 1-step RT-qPCR			
	one rxn		
GoTAq qPCR Master Mix	10.0	) ul	
GoScript RT Mix	0.4	l ul	
Combined Primer/Probe Mix	1.5 ul		
Nuclease-free water	3.1	ul	
	15.0	) ul	
	15.0 ul/sa	mple + 5 ul RNA	
Thermocycle Program			
Reverse Transcriptase	45 °C	15 min	
RT Inactivation	95 °C	2 min	
Denaturation	95 °C	15  sec	45 X
Annealing/Extension *	60 °C	1 min	* Detect FAN

**QPCR** fluorescence readings were measured by a BioRad CFX 96 Real-Time PCR System. The threshold for determining Ct value for each sample is based on the baseline threshold – above background and within the exponential phase of amplification curve.



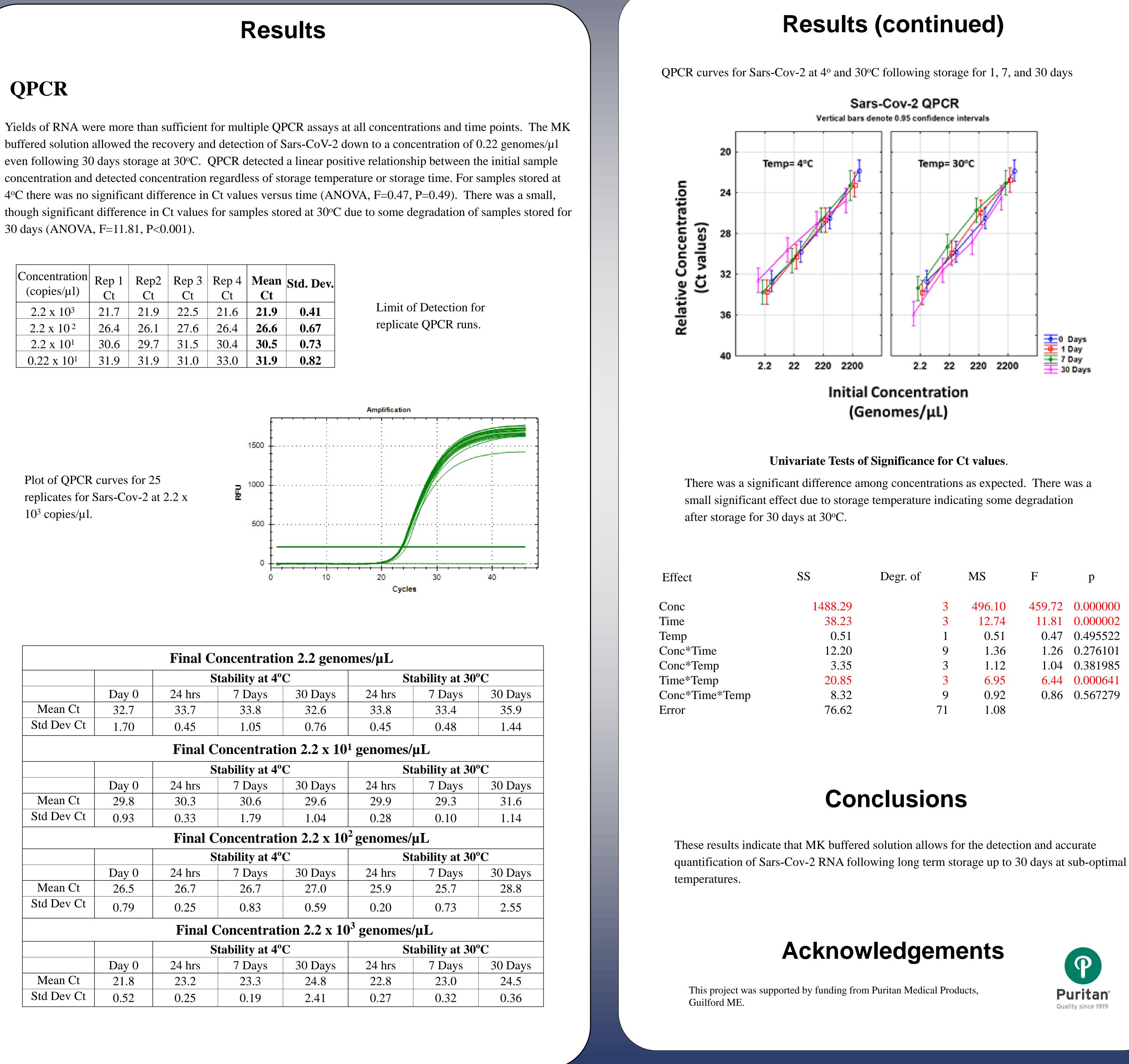
# **Evaluating preservation and quantification of SARS-Cov-2 RNA From a Collection and Transport System for Rapid Point-of-Care Diagnostic Tests**

## **QPCR**

4°C there was no significant difference in Ct values versus time (ANOVA, F=0.47, P=0.49). There was a small, 30 days (ANOVA, F=11.81, P<0.001).

Concentration (copies/µl)	Rep 1 Ct	Rep2 Ct	Rep 3 Ct	Rep 4 Ct	Mean Ct	Std. Dev.
$2.2 \times 10^3$	21.7	21.9	22.5	21.6	21.9	0.41
$2.2 \times 10^{2}$	26.4	26.1	27.6	26.4	26.6	0.67
$2.2 \times 10^{1}$	30.6	29.7	31.5	30.4	30.5	0.73
$0.22 \text{ x } 10^{1}$	31.9	31.9	31.0	33.0	31.9	0.82

Plot of QPCR curves for 25 replicates for Sars-Cov-2 at 2.2 x  $10^3$  copies/µl.



		Final Co	oncentratio	on 2.2 gen	
		Stability at 4°C			
	Day 0	24 hrs	7 Days	30 Days	
Mean Ct	32.7	33.7	33.8	32.6	
Std Dev Ct	1.70	0.45	1.05	0.76	
		<b>Final C</b>	oncentrati	on 2.2 x 1	
		Stability at 4°C			
	Day 0	24 hrs	7 Days	30 Days	
Mean Ct	29.8	30.3	30.6	29.6	
Std Dev Ct	0.93	0.33	1.79	1.04	
		Final C	oncentrati	on 2.2 x 1	
		Stability at 4°C			
	Day 0	24 hrs	7 Days	30 Days	
Mean Ct	26.5	26.7	26.7	27.0	
Std Dev Ct	0.79	0.25	0.83	0.59	
		Final C	Concentrat	ion 2.2 x 1	
		Stability at 4°C			
	Day 0	24 hrs	7 Days	30 Days	
Mean Ct	21.8	23.2	23.3	24.8	
Std Dev Ct	0.52	0.25	0.19	2.41	

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SS	Degr. of	MS	F	р
1488.29	3	496.10	459.72	0.000000
38.23	3	12.74	11.81	0.000002
0.51	1	0.51	0.47	0.495522
12.20	9	1.36	1.26	0.276101
3.35	3	1.12	1.04	0.381985
20.85	3	6.95	6.44	0.000641
8.32	9	0.92	0.86	0.567279
76.62	71	1.08		